

Canada.ca > Health > Health risks and safety > Biosafety and biosecurity

> Pathogen Safety Data Sheets

Pathogen Safety Data Sheets: Infectious Substances – Staphylococcus aureus

PATHOGEN SAFETY DATA SHEET -INFECTIOUS SUBSTANCES

SECTION I - INFECTIOUS AGENT

NAME: Staphylococcus aureus

SYNONYM OR CROSS REFERENCE: MRSA (methicillin-resistant *Staphylococcus aureus*), MSSA (methicillin-susceptive (or sensitive) *Staphylococcus aureus*), VISA (vancomycin-intermediate *Staphylococcus aureus*), hVISA (heteroresistant vancomycin-intermediate *Staphylococcus aureus*), VRSA (vancomycin-resistant *Staphylococcus aureus*), staph infection, staphylococcus infection, impetigo, toxic shock syndrome.

CHARACTERISTICS: *Staphylococcus aureus* are Gram-positive, catalase positive cocci belonging to the **Staphylococcaceae** family ¹, ². They are approximately 0.5-1.5 µm in diameter, nonmotile, non-spore-forming, facultative anaerobes (with the exception of *S. aureus anaerobius*) that usually form in clusters. Many strains produce staphylococcal enterotoxins,

the superantigen toxic shock syndrome toxin (TSST-1), and exfoliative toxins. *Staphylococcus aureus* are part of human flora, and are primarily found in the nose and skin $\frac{3}{2}$.

SECTION II - HAZARD IDENTIFICATION

PATHOGENICITY/TOXICITY: Staphylococcus aureus is an opportunistic pathogen that can cause a variety of self-limiting to life-threatening diseases in humans $\frac{2}{2}$. The bacteria are a leading cause of food poisoning, resulting from the consumption of food contaminated with enterotoxins $\frac{4}{2}$. Staphylococcal food intoxication involves rapid onset of nausea, vomiting, abdominal pain, cramps, and diarrhea $\frac{2}{4}$. Symptoms usually resolve after 24 hours ⁴. Animal bites can result in local infections, cellulitis, erythema, tenderness, mild fever, adenopathy, and lymphangitis (rarely) ⁵. Scalded skin syndrome is caused by exfoliative toxins secreted on the epidermis and mostly affects neonates and young children $\frac{2}{2}$. Other skin conditions caused by Staphylococcal exfoliative toxins include blisters, skin loss, pimples, furuncles, impetigo, folliculitis, abscesses, poor temperature control, fluid loss, and secondary infection ^{2, 4, 6, 7}. S. aureus can also cause necrotizing fasciitis in immunocompromised individuals, although this is very rare ⁸. Necrotizing fasciitis is life-threatening and causes severe morbidity.

Certain strains of *S. aureus* produce the superantigen TSST-1, which is responsible for 75% of toxic shock syndrome (TSS) cases ². The clinical presentation of TSS is severe and acute symptoms include high fever, vascular collapse, vomiting, diarrhea, myalgia, hypotension, erythematous rash, desquamation, and involvement of at least 3 organs ². ⁹. ¹⁰. Mortality is very high and death can occur within 2 hours ⁹. Toxic shock syndrome is associated with vaginal colonization with toxin-producing *S. aureus* during menstruation, complications with staphylococcal infection at other sites, or

complications of surgical procedures $\frac{10}{10}$. Deep infections include endocarditis, peritonitis, necrotizing pneumonia, bacteremia, meningitis, osteomyelitis, septic arthritis, and infections of bones, joints and organs $\frac{2}{7}$.

EPIDEMIOLOGY: Worldwide distribution. *Staphylococcus aureus* is one of the most common causes of skin, soft-tissue, and nosocomial infection $\frac{7}{2}$. Rates of infection in community settings are increasing $\frac{7}{2}$, $\frac{11}{2}$. Residents of nursing homes are also at an increased risk of acquiring MRSA $\frac{12}{2}$. Around 20% of individuals are persistent carriers of *Staphylococcus aureus*, about 60% are intermittent carriers, and approximately 20% rarely carry it $\frac{3}{2}$. Children are more likely to be persistent carriers of the bacteria $\frac{3}{2}$. Young women are at a higher risk for toxic shock syndrome $\frac{10}{2}$.

HOST RANGE: Humans, wild and domestic animals, including cows $\frac{13}{2}$.

INFECTIOUS DOSE: At least 100,000 organisms in humans ¹⁴.

MODE OF TRANSMISSION: Ingestion of food containing enterotoxins $\frac{4}{5}$. Vertical transmission during vaginal delivery is uncommon $\frac{15}{5}$. Person-toperson transmission occurs through contact with a purulent lesion or with a carrier $\frac{3}{5}$. Unsanitary conditions and crowded community settings increase exposure to *S. aureus* $\frac{16}{5}$. Infection may be spread from person-toperson through health care workers or patients $\frac{3}{5}$. Nasal colonization can lead to auto-infection $\frac{17}{5}$.

INCUBATION PERIOD: Onset of symptoms after consuming contaminated food is usually 30 minutes to 8 hours $\frac{4}{2}$. Colonies of S. aureus can be carried for an undetermined amount of time; some individuals may carry it chronically, and some may carry it intermittently $\frac{3}{2}$.

COMMUNICABILITY: Communicable period is as long as a purulent lesion is present or carrier state persists.

SECTION III - DISSEMINATION

RESERVOIR: *Staphylococcus aureus* is found in humans in the nose, groin, axillae, perineal area (males), mucous membranes, the mouth, mammary glands, hair, and the intestinal, genitourinary and upper respiratory tracts $\frac{2}{4}$, $\frac{4}{18}$. Many animals act as reservoirs, particularly cows with infected udders $\frac{13}{18}$.

ZOONOSIS: Yes, through direct or indirect contact with an infected animal <u>⁵</u>.

VECTORS: None.

SECTION IV - STABILITY AND VIABILITY

DRUG SUSCEPTIBILITY: Antibiotics such as cloxacillin and cephalexin are commonly used to treat staph infections $\frac{19}{20}$. Vancomycin which is administered intravenously is used to treat MRSA $\frac{20}{20}$.

DRUG RESISTANCE: Many strains of Staphylococcus aureus have increasing resistance to multiple antibiotic classes $\frac{6}{2}$. Methicillin resistant strains are common causes of nosocomial infection $\frac{21}{2}$. Increasing resistance to vancomycin is being documented in many hospitals $\frac{6}{2}$.

SUSCEPTIBILITY TO DISINFECTANTS: Susceptible to 70% ethanol, clorhexidine, 1% sodium hypochlorite, 2% glutaraldehyde, 0.25% benzalkonium chloride, and formaldehyde ^{12, 22, 23}.

PHYSICAL INACTIVATION: *Staphylococcus aureus* can grow in a pH of 4.2 to 9.3 and in salt concentrations of up to 15% $\frac{4}{2}$. Enterotoxins are resistant to temperatures that would destroy the bacilli $\frac{4}{2}$. Sensitive to dry heat treatment of 160-170°C for at least an hour, but not to moist heat treatment $\frac{24}{2}$.

SURVIVAL OUTSIDE HOST: Survives on carcasses and organs (up to 42 days), floors (less than 7 days), glass (46 hours), sunlight (17 hours), UV (7 hours), meat products (60 days), coins (up to 7 days), skin (30 minutes to 38 days) (citation needed). Depending on colony size, *S. aureus* can survive on fabrics from days to months $\frac{25}{25}$.

SECTION V - FIRST AID / MEDICAL

SURVEILLANCE: Monitor for symptoms. In outbreak settings, food poisoning can be diagnosed on clinical grounds with food cultured for *S*. *aureus* 2 . Toxic shock syndrome can be indicated with a clinical diagnosis and isolation of *S*. *aureus* strain, TSST-1, or enterotoxins B or C. This can be achieved using ELISA, reverse passive latex agglutination, or PCR. Scalded skin syndrome can be diagnosed clinically, with presence of Nikolsky's sign and identification of *S*. *aureus* retrieved from the infection site. Bacteremia and deep site infections are confirmed with direct microscopic examination of clinical specimen.

Note: All diagnostic methods are not necessarily available in all countries.

FIRST AID/TREATMENT: Treatment of abscesses usually does not need antibiotic therapy; appropriate drainage is usually sufficient $\frac{6}{2}$. Proper antibiotic therapy is required for more serious infections.

IMMUNIZATION: None ².

PROPHYLAXIS: Elimination of nasal carriage by using topical mupirocin also eliminates hand carriage $\frac{3}{2}$.

SECTION VI - LABORATORY HAZARDS

LABORATORY-ACQUIRED INFECTIONS: 29 reported cases as of 1973, with 1 death $\frac{26}{2}$.

SOURCE/SPECIMENS: Infective stages may be present in CSF, joint aspirates, blood, abscesses, aerosols, faeces, and urine $\frac{2}{2}$, $\frac{4}{2}$, $\frac{6}{2}$, $\frac{18}{2}$.

PRIMARY HAZARDS: Trauma of cutaneous barrier, parenteral inoculation, direct implantation of medical devices (i.e. indwelling catheters and IVs), ingestion of infected material, and contact with aerosols $\frac{2}{2}$, $\frac{4}{2}$, $\frac{18}{2}$.

SPECIAL HAZARDS: Contaminated request forms that have been wrapped around specimen containers $\frac{21}{2}$. Direct contact with open cuts and lesions of skin.

SECTION VII – EXPOSURE CONTROLS / PERSONAL PROTECTION

RISK GROUP CLASSIFICATION: Risk Group 2 27.

CONTAINMENT REQUIREMENTS: Containment Level 2 facilities, equipment, and operational practices for work involving infectious or potentially infectious materials, animals, or cultures.

PROTECTIVE CLOTHING: Lab coat. Gloves when direct skin contact with infected materials or animals is unavoidable. Eye protection must be used where there is a known or potential risk of exposure to splashes $\frac{28}{28}$.

OTHER PRECAUTIONS: All procedures that may produce aerosols, or involve high concentrations or large volumes should be conducted in a biological safety cabinet (BSC). The use of needles, syringes, and other sharp objects should be strictly limited. Additional precautions should be considered with work involving animals or large scale activities $\frac{28}{28}$.

SECTION VIII - HANDLING AND STORAGE

SPILLS: Allow aerosols to settle and, wearing protective clothing, gently cover spill with paper towels and apply an appropriate disinfectant, starting at the perimeter and working towards the centre. Allow sufficient contact time before clean up.

DISPOSAL: Decontaminate all wastes that contain or have come in contact with the infectious organism before disposing by autoclave, chemical disinfection, gamma irradiation, or incineration.

STORAGE: The infectious agent should be stored in leak-proof containers that are appropriately labelled.

SECTION IX - REGULATORY AND OTHER INFORMATION

REGULATORY INFORMATION: The import, transport, and use of pathogens in Canada is regulated under many regulatory bodies, including the Public Health Agency of Canada, Health Canada, Canadian Food Inspection Agency, Environment Canada, and Transport Canada. Users are responsible for ensuring they are compliant with all relevant acts, regulations, guidelines, and standards.

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PREPARED BY: Pathogen Regulation Directorate, Public Health Agency of Canada

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REFERENCES:

- <u>1</u> Becker, K., Harmsen, D., Mellmann, A., Meier, C., Schumann, P., Peters, G., & von Eiff, C. (2004). Development and evaluation of a quality-controlled ribosomal sequence database for 16S ribosomal DNA-based identification of Staphylococcus species. *Journal of Clinical Microbiology, 42*(11), 4988-4995. doi:10.1128/JCM.42.11.4988-4995.2004
- Murray, P. R., Baron, E. J., Jorgensen, J. H., Landry, M. L., Pfaller, M. A., & Yolken, R. H. (Eds.). (2003). *Manual of Clinical Microbiology* (8th ed.). Herdon, VA, United States of America: American Society for Microbiology.
- Skiuytmans, J., van Belkum, A., & Verbrugh, H. (1997). Nasal carriage of Staphylococcus aureus: epidemiology, underlying mechanisms, and associated risks. *Clinical Microbiology Reviews*, *10*(3), 505-520.
- Le Loir, Y., Baron, F., & Gautier, M. (2003). Staphylococcus aureus and food poisoning. *Genetics and Molecular Research : GMR, 2*(1), 63-76.
- 5 Goldstein, E. J., Citron, D. M., Wield, B., Blachman, U., Sutter, V. L., Miller, T. A., & Finegold, S. M. (1978). Bacteriology of human and animal bite wounds. *Journal of Clinical Microbiology, 8*(6), 667.

- Eisenstein, B. I. (2008). Treatment challenges in the management of complicated skin and soft-tissue infections. *Clinical Microbiology and Infection : The Official Publication of the European Society of Clinical Microbiology and Infectious Diseases, 14 Suppl 2*, 17-25. doi:10.1111/j.1469-0691.2008.01922.x
- Fridkin, S. K., Hageman, J. C., Morrison, M., Sanza, L. T., Como-Sabetti, K., Jernigan, J. A., Harriman, K., Harrison, L. H., Lynfield, R., & Farley, M. M. (2005). Methicillin-resistant Staphylococcus aureus disease in three communities. *The New England Journal of Medicine*, 352(14), 1436.
- Miller, L. G., Perdreau-Remington, F., Rieg, G., Mehdi, S., Perlroth, J., Bayer, A. S., Tang, A. W., Phung, T. O., & Spellberg, B. (2005).
 Necrotizing fasciitis caused by community-associated methicillin-resistant Staphylococcus aureus in Los Angeles. *The New England Journal of Medicine*, 352(14), 1445.
- Chen, C., Tang, J., Dong, W., Wang, C., Feng, Y., Wang, J., Zheng, F., Pan, X., Liu, D., Li, M., Song, Y., Zhu, X., Sun, H., Feng, T., Guo, Z., Ju, A., Ge, J., Dong, Y., Sun, W., Jiang, Y., Wang, J., Yan, J., Yang, H., Wang, X., Gao, G. F., Yang, R., Wang, J., & Yu, J. (2007). A glimpse of streptococcal toxic shock syndrome from comparative genomics of S. suis 2 Chinese isolates. *PloS One, 2*(3), e315. doi:10.1371/journal.pone.0000315
- Parsonnet, J., Hansmann, M. A., Delaney, M. L., Modern, P. A., DuBois, A. M., Wieland-Alter, W., Wissemann, K. W., Wild, J. E., Jones, M. B., & Seymour, J. L. (2005). Prevalence of toxic shock syndrome toxin 1-producing Staphylococcus aureus and the presence of antibodies to this superantigen in menstruating women. *Journal of Clinical Microbiology*, *43*(9), 4628.

- David, M. D., Kearns, A. M., Gossain, S., Ganner, M., & Holmes, A.
 (2006). Community-associated meticillin-resistant Staphylococcus aureus: nosocomial transmission in a neonatal unit. *The Journal of Hospital Infection, 64*(3), 244-250. doi:10.1016/j.jhin.2006.06.022
- Hughes, C. M., Smith, M. B., & Tunney, M. M. (2008). Infection control strategies for preventing the transmission of meticillin-resistant Staphylococcus aureus (MRSA) in nursing homes for older people. *Cochrane Database of Systematic Reviews (Online), (1)* (1), CD006354. doi:10.1002/14651858.CD006354.pub2
- Fitzgerald, J. R., Sturdevant, D. E., Mackie, S. M., Gill, S. R., & Musser, J. M. (2001). Evolutionary genomics of Staphylococcus aureus: insights into the origin of methicillin-resistant strains and the toxic shock syndrome epidemic. *Proceedings of the National Academy of Sciences of the United States of America, 98*(15), 8821-8826. doi:10.1073/pnas.161098098
- <u>14</u> Schmid-Hempel, P., & Frank, S. A. (2007). Pathogenesis, virulence, and infective dose. *PLoS Pathogens, 3*(10)
- Reusch, M., Ghosh, P., Ham, C., Klotchko, A., Singapuri, S., &
 Everett, G. (2008). Prevalence of MRSA colonization in peripartum mothers and their newborn infants. *Scandinavian Journal of Infectious Diseases, 40*(8), 667-671.
- Stevens, A. M., Hennessy, T., Baggett, H. C., Bruden, D., Parks, D., & Klejka, J. (2010). Methicillin-Resistant Staphylococcus aureus Carriage and Risk Factors for Skin Infections, Southwestern Alaska, USA. *Emerging Infectious Diseases, 16*(5), 797.

- van Belkum, A., Emonts, M., Wertheim, H., de Jongh, C., Nouwen, J., Bartels, H., Cole, A., Cole, A., Hermans, P., Boelens, H., Toom, N. L., Snijders, S., Verbrugh, H., & van Leeuwen, W. (2007). The role of human innate immune factors in nasal colonization by Staphylococcus aureus. *Microbes and Infection / Institut Pasteur, 9*(12-13), 1471-1477. doi:10.1016/j.micinf.2007.08.003
- <u>18</u> Spendlove, J. C., & Fannin, K. F. (1983). Source, significance, and control of indoor microbial aerosols: human health aspects. *Public Health Reports, 98*(3), 229.
- <u>19</u> Sharma, S., & Verma, K. K. (2001). Skin and soft tissue infection. *Indian Journal of Pediatrics, 68 Suppl 3*, S46-50.
- Hansra, N. K., & Shinkai, K. (2011). Cutaneous communityacquired and hospital-acquired methicillin-resistant
 Staphylococcus aureus. Dermatologic Therapy, 24(2), 263-272. doi:10.1111/j.1529-8019.2011.01402.x; 10.1111/j.1529-8019.2011.01402.x
- 21 Collins, C. H., & Kennedy, D. A. (1999). Laboratory-acquired infections. *Laboratory-acquired Infections: History, incidence, causes and prevention.* (4th ed., pp. 234). Oxford, UK: Butterworth Heinemann.
- Sinha, B., François, P. P., Nüße, O., Foti, M., Hartford, O. M.,
 Vaudaux, P., Foster, T. J., Lew, D. P., Herrmann, M., & Krause, K. H. (1999). Fibronectin-binding protein acts as Staphylococcus aureus invasin via fibronectin bridging to integrin α5β1. *Cellular Microbiology*, *1*(2), 101-117.

- 23 Egusa, H., Watamoto, T., Matsumoto, T., Abe, K., Kobayashi, M., Akashi, Y., & Yatani, H. (2008). Clinical evaluation of the efficacy of removing microorganisms to disinfect patient-derived dental impressions. *The International Journal of Prosthodontics, 21*(6), 531-538.
- Moesby, L., Hansen, E. W., Christensen, J. D., Hoyer, C. H., Juhl, G. L., & Olsen, H. B. (2005). Dry and moist heat sterilisation cannot inactivate pyrogenicity of Gram positive microorganisms.
 European Journal of Pharmaceutical Sciences : Official Journal of the European Federation for Pharmaceutical Sciences, 26(3-4), 318-323. doi:10.1016/j.ejps.2005.07.003
- 25 Neely, A. N., & Maley, M. P. (2000). Survival of enterococci and staphylococci on hospital fabrics and plastic. *Journal of Clinical Microbiology*, *38*(2), 724-726.
- <u>26</u> Pike, R. M. (1976). Laboratory-associated infections: summary and analysis of 3921 cases. *Health Laboratory Science*, *13*(2), 105-114.
- Human Pathogens and Toxins Act. S.C. 2009, c. 24. Government of Canada, Second Session, Fortieth Parliament, 57-58 Elizabeth II, 2009, (2009).
- <u>28</u> Public Health Agency of Canada. (2004). In Best M., Graham M. L.,
 Leitner R., Ouellette M. and Ugwu K. (Eds.), *Laboratory Biosafety Guidelines* (3rd ed.). Canada: Public Health Agency of Canada.

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